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Gaussian process modeling of bioprocesses: application to chinese ovary hamster cells cultivated in bioreactors

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November 25, 2021

• Monoclonal antibodies (mAb): inhibition of viral infection.



Figure: Scheme of mAb against COVID-19<sup>1</sup>

<sup>1</sup>Source: website of National Institutes of Health (https://www.nih.gov/)

• In industry, mAb is produced by Chinese Hamster Ovary (CHO) cells.



• Chinese Hamster Ovary (CHO) cells cultivated in large-scale bioreactors.



Figure: Picture of a bioreactor<sup>2</sup>

 $^{2} https://www.engr.colostate.edu/CBE101/topics/bioreactors.html$ 



Figure: Scheme of a bioreactor<sup>3</sup>

<sup>3</sup>https://en.wikipedia.org/wiki/Bioreactor/media/File:Bioreactor\_principle.svg

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• Producing mAb is expensive (and so are the treatments)!

• Ronapreve approved by EU commission against COVID-19 at 1.700 EUR<sup>4</sup>

• Goal: maximization of the yield of mAb produced by CHO cells.

• Which variable should we change for the maximization?

<sup>4</sup>https://www.leparisien.fr/societe/sante/ronapreve-regkirona-ce-que-lon-sait-des-deux-traitements-approuves-par-lema-11-11-2021-U64NC7CAQJE2XGVSVW2QF7FIBM.php



Figure: Scheme of a bioreactor

# How to optimize the feed medium?

Optimize experimentally? No, too long and expensive!

Solution: model-based optimization.

• What to model? Kinetic evolution of the concentrations in the bioreactor w.r.t. variations of the feed medium concentrations.



# Some first principles?

• The model should satisfy some first principles such as mass-balance equation

Evolution \_ What the cells \_ What the feed \_ What the effluent \_ takes out \_ takes out \_ takes out \_ takes out \_ takes \_ take

• With mathematics (x = concentration vector)

$$\frac{dx}{dt}(t) = q(x(t)) + F_{in}(t)x_{feed}(t) - F_{out}(t)x(t)$$

where

- q: uptake/secretion rate (quantity of metabolites consumed/produced by the cells during one day)
- $F_{in}$ : flow rate of feed medium.
- $F_{out}$ : flow rate of effluent.
- $x_{feed}$ : concentration of metabolites in the feed medium.

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- q: uptake/secretion rate (quantity of metabolites consumed/produced by the cells during one day)
- $F_{in}$ : flow rate of feed medium.
- $F_{out}$ : flow rate of effluent.
- $x_{feed}$ : concentration of metabolites in the medium feed.

 $\bullet$  The vector  $q(\boldsymbol{x}(t))$  describes the kinetics of the chemical reactions inside the cells.

- $\bullet$  To model  $q(\boldsymbol{x}(t)),$  we need 3 ingredients
  - Connections between the metabolites (metabolic network).
  - Stoichiometric coefficients of the reactions (stoichiometric matrix)
  - Rate (speed) of all the chemical reactions (kinetic expression)

# Ingredient 1: metabolic network



## REAL metabolic network



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# Ingredient 2: Stoichiometric matrix

- Stoichiometric coefficients:
  - Reaction 1:  $S_1 \rightarrow 2C_1$
  - Reaction 2:  $S_2 \rightarrow C_1 + C_2$
  - Reaction 3:  $C_1 + 3C_2 \rightarrow P$

	Γ	Reaction 1	Reaction 2	Reaction 3
	$S_1$	-1	0	0
۸	$S_2$	0	-1	0
A =	$C_1$	2	1	-1
	$C_2$	0	1	-3
	P	0	0	1

• Then,

$$q(x) = \mathbf{A} y(x)$$

# Ingredient 3: Kinetic expression of the rates

• **Parametric** expression for the rate vector y(x).

• For biochemical reactions: Monod kinetics

Rate of Reaction i = 
$$\prod_{j=1}^{n}$$
 Modulation function $(x_j, \eta_{ij})$ 

• 4 types of modulation functions:

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# Ingredient 3: Kinetic expression of the rates



• Final mass balance-equation with kinetics

$$\frac{dx}{dt}(t) = \mathbf{A}y(x(t), \boldsymbol{\eta}) + F_{in}(t)x_{feed}(t) - F_{out}(t)x(t)$$

• Goal: identify  $\eta$ .

• Everything is measured/known except  $y(x(t), \pmb{\eta}).$  We can estimate the data of the rates!

# From dynamic to static map modeling

• Data:  $\{y(t), x(t)\}_{t=1}^N.$ 

• The identification problem for the *i*-th rate is

$$\eta_i^* = \arg \min \sum_{t=1}^N ||y_i(t) - y_{m,i}(x(t), \eta_i)||^2$$

where

$$y_{m,i}(x(t),\eta_i) = \prod_{j=1}^n \frac{x_j(t)}{(x_j(t) + \theta_{ij})(1 + \mu_{ij}x_j(t))}$$

• We have transformed the nonlinear dynamic identification problem into a nonlinear static map identification.

## Design of a kernel for Gausian process regression of Monod functions

• Focus on one rate.

• The identification problem is

$$\eta^* = \arg\,\min\sum_{t=1}^N ||y(t)-y_m(x(t),\eta)||^2$$

where

$$y_m(x(t),\eta) = \prod_{j=1}^n \frac{x_j(t)}{(x_j(t) + \theta_j)(1 + \mu_j x_j(t))}$$

• Local minima: we need a good initialization of the parameters.

• Main idea: perform a GP regression of **each** modulation function as intermediate modeling method for the initialization.

• Assume additive noise:

$$y(t) = \prod_{j=1}^{n} h_j(x_j(t), \eta_j) + \epsilon(t)$$

where  $\epsilon$  is a white Gaussian noise of variance  $\sigma_e^2.$ 

• Each function  $h_i$  is modeled as a zero-mean Gaussian process

$$h_j \sim \mathcal{GP}(0, k_j)$$

• We need an appropriate kernel function for the modeling of the 4 types of Monod functions.

• Kernel proposed in the literature<sup>5</sup>

$$k_j(x_j(t), x_j(t'), \Theta_j) = \gamma_j \left(\frac{x_j(t)}{x_j(t')}\right)^{-\delta_j \log\left(\frac{x_j(t)}{x_j(t')}\right)}$$

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<sup>&</sup>lt;sup>5</sup>Wang, M., Risuleo, R. S., Jacobsen, E. W., Chotteau, V., Hjalmarsson, H. (2020). Identification of nonlinear kinetics of macroscopic bio-reactions using multilinear Gaussian processes. Computers Chemical Engineering, 133, 106671

# Kernel in the literature



Figure: Some examples of posterior means<sup>6</sup>

<sup>6</sup>Wang, M., Risuleo, R. S., Jacobsen, E. W., Chotteau, V., Hjalmarsson, H. (2020). Identification of nonlinear kinetics of macroscopic bio-reactions using multilinear Gaussian processes. Computers Chemical Engineering, 133, 106671

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# Kernel in the literature

• However, it can yield inaccurate estimates for small data set and/or poorly distributed in the input space ( $\approx$  bioreactor data).

 $\bullet$  Example with y=x/(x+10) with N=5 noiseless data randomly chosen in the interval  $\left[80,130\right]$ 



• What to do?

• Idea 1: experiment design (but it is costly!).

• Idea 2: incorporate priors in the kernel design.

#### Research problem

Design better-tailored kernel function for the modeling of the Monod functions  $h_j$ .

• Idea: incorporate the **structure** of the Monod functions in the design.

# Design of a better kernel: activation

• Consider again a simple activation function for the true system

$$y(x(t)) = h_{act}(x(t), \theta) + \epsilon$$

where  $h_{act}(x(t)) = x(t)/(x(t) + \theta)$ .

 $\bullet$  The covariance between two output data  $y(\boldsymbol{x}(t))$  and  $y(\boldsymbol{x}(t'))$  is equal to

$$E[y(x(t))y(x(t'))] = h_{act}(x(t),\theta)h_{act}(x(t'),\theta) + E[\epsilon(t)\epsilon(t')]$$

 $\bullet$  Ideal kernel function for modeling of  $h_{act}$  is then

$$k_{act,ideal}(x(t), x(t'), \theta) = h_{act}(x(t), \theta) h_{act}(x(t'), \theta)$$

• But (i) this kernel is invalid (covariance matrix never positive definite) and (ii) it depends on the unknown  $\theta$ 

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# Design of a better kernel: activation

• Idea: integrate w.r.t.  $\theta$  between two bounds  $\theta^-$  and  $\theta^+!$ 

$$\begin{aligned} k_{act}(x,x') &= \int_{\theta^{-}}^{\theta^{+}} h_{act}(x,\theta) h_{act}(x',\theta) d\theta \\ &= \begin{cases} x^{2} \left(\frac{1}{\theta^{-}+x} - \frac{1}{\theta^{+}+x}\right) & \text{if } x = x' \\ \frac{xx'}{x-x'} \log \left(\frac{\theta^{+}+x'}{\theta^{+}+x} \cdot \frac{\theta^{-}+x}{\theta^{-}+x'}\right) & \text{elsewhere} \end{cases} \end{aligned}$$

• We obtain a valid kernel constructed directly from activation function (prior added in the design)!

• But, two additional hyperparameters  $(\theta^- \text{ and } \theta^+)!$ 

• How to choose  $\theta^-$  and  $\theta^+$ ?

• If no prior on kinetic parameters  $\theta$ , we choose "wide bounds" but still biologically realistic (e.g.,  $\theta^- = 0.01$  and  $\theta^+ = 100$ ).

• However, if we have priors on  $\theta$  in the form of uncertainty intervals

$$\theta \in [\Theta^-, \Theta^+]$$

then we choose  $\theta^- = \Theta^-$  and  $\theta^+ = \Theta^+$ .

# Design of a better kernel: activation

• Back to the example with bounds  $\theta^- = 0.01$  and  $\theta^+ = 100$ .



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# Design of a better kernel: activation

• Assume that we know that  $\theta \in [5, 20]$ . Chosen bounds:  $\theta^- = 5$ ,  $\theta^+ = 20$ .



# Design of a better kernel: inhibition

Inhibition function

$$h_{inh}(x) = 1/(\mu x + 1)$$

• We can similarly construct a kernel with integration:

$$\begin{aligned} k_{inh}(x,x') &= \int_{\mu^{-}}^{\mu^{+}} h_{inh}(x,\mu) h_{inh}(x',\mu) d\mu \\ &= \begin{cases} &\frac{1}{x} \left( \frac{1}{\mu^{-}x+1} - \frac{1}{\mu^{+}x+1} \right) & \text{if } x = x' \\ &\frac{1}{x-x'} \log \left( \frac{\mu^{+}x+1}{\mu^{+}x'+1} \cdot \frac{\mu^{-}x+1}{\mu^{-}x'+1} \right) & \text{elsewhere} \end{cases} \end{aligned}$$

• Double-component

$$h_{dc}(x) = h_{act}(x,\theta)h_{inh}(x,\mu)$$

• New kernel:

$$k_{dc}(x, x') = k_{act}(x, x')k_{inh}(x, x')$$

# Design of a better kernel: neutral effect

• Neutral effect

$$h_{ne}(x) = 1$$

• We should consider a mean function and a zero covariance.

• Instead of this, we approximate for neutral function by

$$h_{ne}(x) = 1 + \varepsilon x$$

with  $\varepsilon \ll 1$  (10<sup>-3</sup>).

• Then,  $k_{ne}(x, x') = (1 + \varepsilon x)(1 + \varepsilon x').$ 

• We have a kernel for each type of kinetics. For each modulation function, we have to select the best kernel.

- BUT, we do not know the type of kinetic beforehand.
- Final kernel:

$$k_j(x_j, x_j') = \frac{\beta_{act,j}k_{act}(x_j, x_j') + \beta_{inh,j}k_{inh}(x_j, x_j') + \frac{\beta_{dc,j}k_{dc}(x_j, x_j') + \beta_{ne,j}k_{ne}(x_j, x_j')}{\beta_{ne,j}k_{ne}(x_j, x_j') + \beta_{ne,j}k_{ne}(x_j, x_j')}$$

• By tuning the hyperparameters  $\beta_j = (\beta_{act,j}, \beta_{inh,j}, \beta_{dc,j}, \beta_{ne,j})^T$ , we can select different types of kinetics.

• With  $\beta_j = (1, 0, 0, 0)^T$ , we will consider activation functions for  $h_j$ .

# Tuning the hyperparameters with Empirical Bayes

# Hyperparameter estimation

- With n different metabolites, we have n hyperparameter vectors  $\beta_j$  to estimate.
- We have data

$$\boldsymbol{y} = \begin{pmatrix} y(1) \\ \vdots \\ y(N) \end{pmatrix}$$

• Empirical Bayes: we want to estimate the hyperparameter vector  $\beta = (\beta_1^T, \cdots, \beta_n)^T$  such that we maximize the likelihood  $p(y|\beta)$ .

• BUT

$$y(t) = \prod_{j=1}^{n} h_j(x_j(t)) + \epsilon(t)$$
$$h_j \sim \mathcal{GP}(0, k_j(\beta_j))$$

ightarrow Likelihood intractable!

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• How to circumvent the problem?

• Solution: introduce latent variables  $h_j = \{h_j(t)\}_{t=1}^N$  for all  $j = 1, \dots, n$  ( $\rightarrow N \times n$  latent variables).

• How to estimate? Iterative estimation of the hyperparameters and latent variables as follows

• From  $\beta^{(k)}$ , we sample the latent variables  $h_j$  from the posterior  $p(h_1, \cdots, h_n | \boldsymbol{y}, \beta^{(k)}) \Longrightarrow \hat{h}_j^{(k)}$ . • From  $\hat{h}_j^{(k)}$ , we estimate  $\beta$  which maximizes  $p(\boldsymbol{h}^{(k)} | \beta) \Longrightarrow \beta^{(k+1)}$ 

 $\blacksquare \ k \to k+1$ 

• Expectation maximization + sampling (heuristic convergence to global optimum for  $\beta$ ).

- How to sample from joint posterior  $p(\boldsymbol{h}_1, \cdots, \boldsymbol{h}_n | \boldsymbol{y}, \beta^{(k)})$ ?
- Solution: Gibbs sampling, i.e., iterative sampling of the conditional posterior distributions.

$$\widehat{\boldsymbol{h}}_1 \to \widehat{\boldsymbol{h}}_2 \to \cdots \to \widehat{\boldsymbol{h}}_n \to \widehat{\boldsymbol{h}}_1 \to \cdots$$

• Hyperparameter optimization?

Solve

$$\beta_j^* = \arg \max_{\beta_j} -\log \det(K(\beta_j)) - \hat{\boldsymbol{h}}_j^T K^{-1}(\beta_j) \hat{\boldsymbol{h}}_j$$
(1)

#### where

$$K(\beta_j) = \beta_{act,j} K_{act} + \beta_{inh,j} K_{inh} + \beta_{dc,j} K_{dc} + \beta_{ne,j} K_{ne}$$
(2)

- Nonconvex optimization:
  - Bruteforce optimization.
  - Combinatorial optomization.

# Numerical example

# Numerical example

• Toy example with n=6 metabolites and  $\sigma_{\epsilon}^2=10^{-3}$ .

$$y = h_1 \times h_2 \times h_3 \times h_4 \times h_5 \times h_6 + \epsilon$$

Modulation function	Type of kinetic	$ heta_j$	$\mu_j$
$h_1$	Activation	8.01	-
$h_2$	Neutral	-	-
$h_3$	Inhibition	-	2.27
$h_4$	Double Component	6.81	0.82
$h_5$	Activation	0.67	_
$h_6$	Inhibition	-	1.8

Table: True type of kinetics and parameters for the 6 modulation functions  $h_i$ .

• N = 30 concentration data chosen randomly in interval [0, 10].

# Numerical example



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# Real-life data with 4 metabolites in kinetic modeling

• Black: data, Blue: kernel from literature, Red and magenta: new kernel













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# Conclusion and possible extension

• Design of a better-tailored kernel for the modeling of Monod functions.

• Better results than kernel in the literature.

• Main idea: incorporate the structure of the functions to be modeled in the kernel design.

• Test on real-life data.

• First possible generalization of the study

$$\int_{\theta^{-}}^{\theta^{+}} h_{act}(x,\theta) h_{act}(x',\theta) d\theta \to \int_{-\infty}^{\infty} h_{act}(x,\theta) h_{act}(x',\theta) p(\theta) d\theta$$

• Second possible extension: assume that we have a static map to be modeled of this form

$$y(x) = \sum_{j=1}^{n} \phi_j(x, \theta_j) + \epsilon$$

where  $\phi_j(x, \theta_j)$  are parametrized nonlinear basis functions (structure known, parameter  $\theta_j$  unknown).

- $\bullet$  Nonlinear least-square optimization  $\rightarrow$  local minimum.
- Idea: GP regression with a kernel equal to a linear combination of kernels designed as

$$k_j(x, x') = \int_{-\infty}^{\infty} \phi_j(x, \theta_j) \phi_j(x', \theta_j) p(\theta_j) d\theta_j$$

# Thank you for your attention!